

Sondas de ácidos nucleicos

Usos:

Para la detección de ácidos nucleicos: Southern-blot, Northern-blot, RNase protection, Dot-blot, Colony-blot etc

Como reactivo: Ej. Para estudiar proteínas de unión ácidos nucleicos, para determinar estructura de RNAs etc.

Métodos para generar sondas:

Nick translation.

Random primers: DNA o RNA

Oligo dT

PCR: marcar primers o producto de PCR.

Primer extension

Transcripción in vitro.

Forforilación 5'.

Transferasa terminal.

Actividad específica: Cantidad de marca en relación a la masa total de sonda.

Southern-Blot



Professor Sir Edwin Southern
University of Oxford

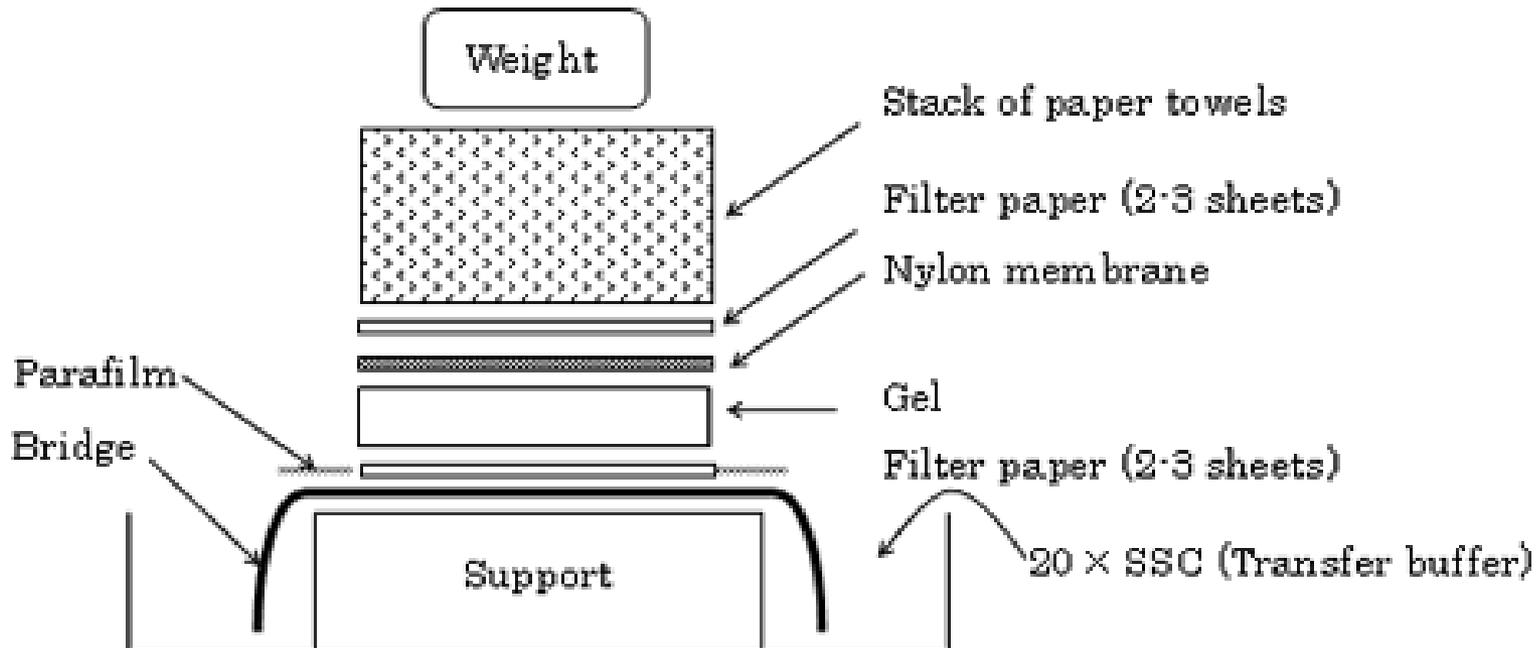
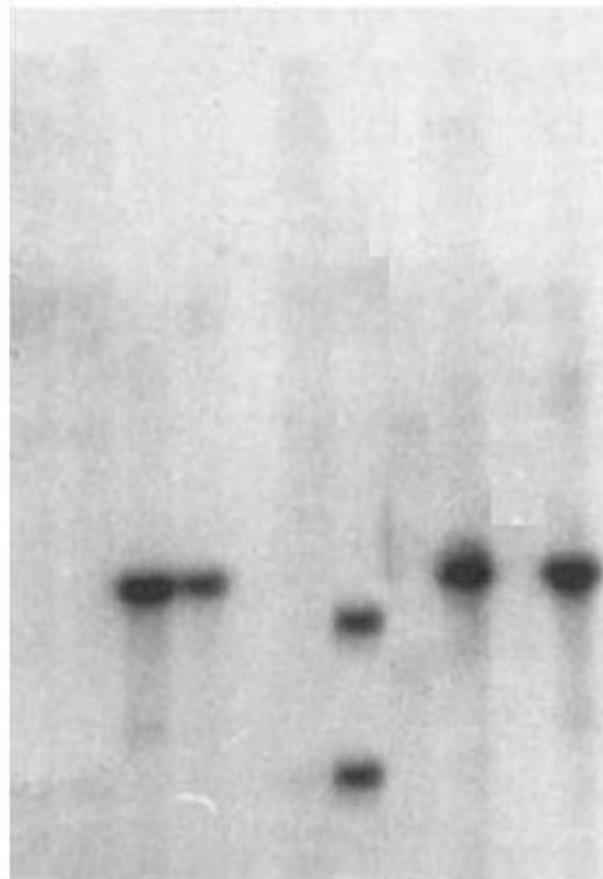
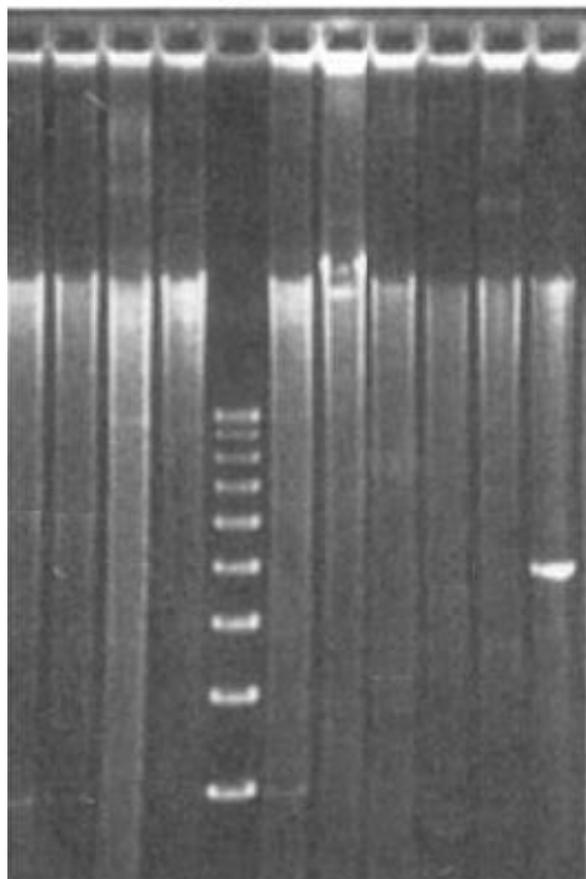
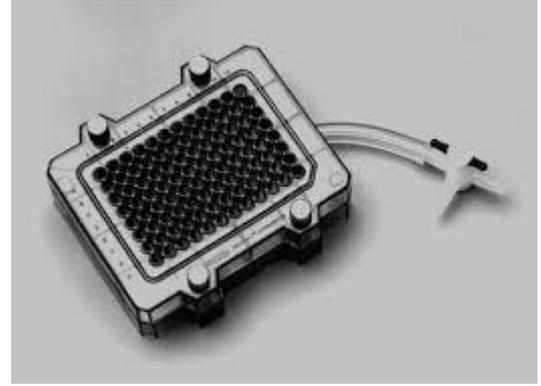
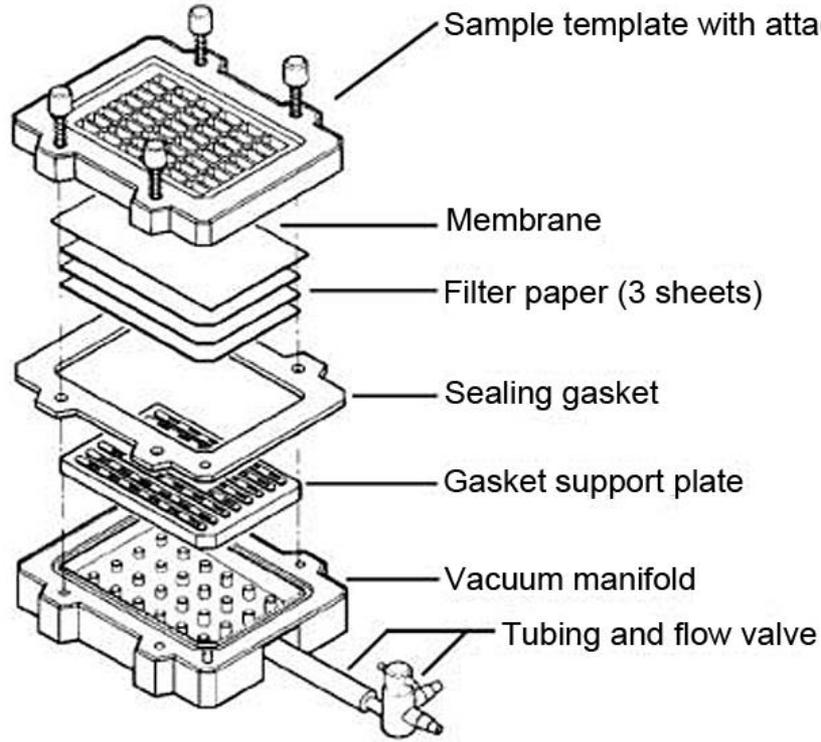
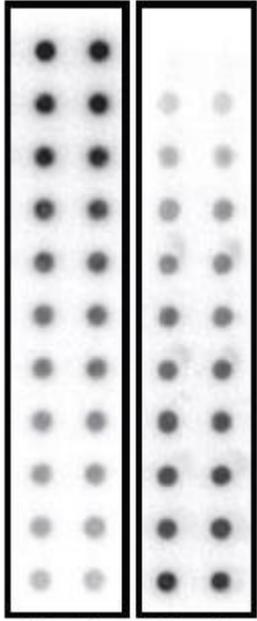


Fig. 1 Transfer Apparatus

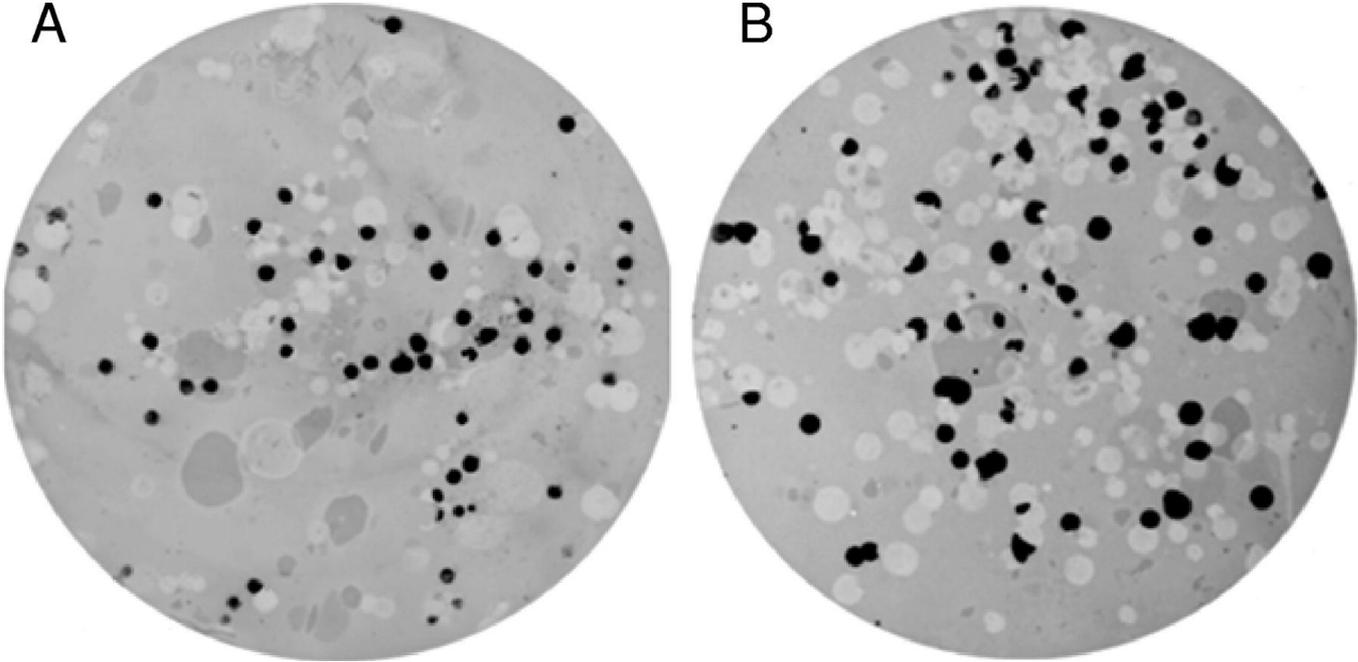
1 2 3 4 (L) 5 6 7 8 9 10 1 2 3 4 (L) 5 6 7 8 9 10



Dot-Blot



Colony-Blot



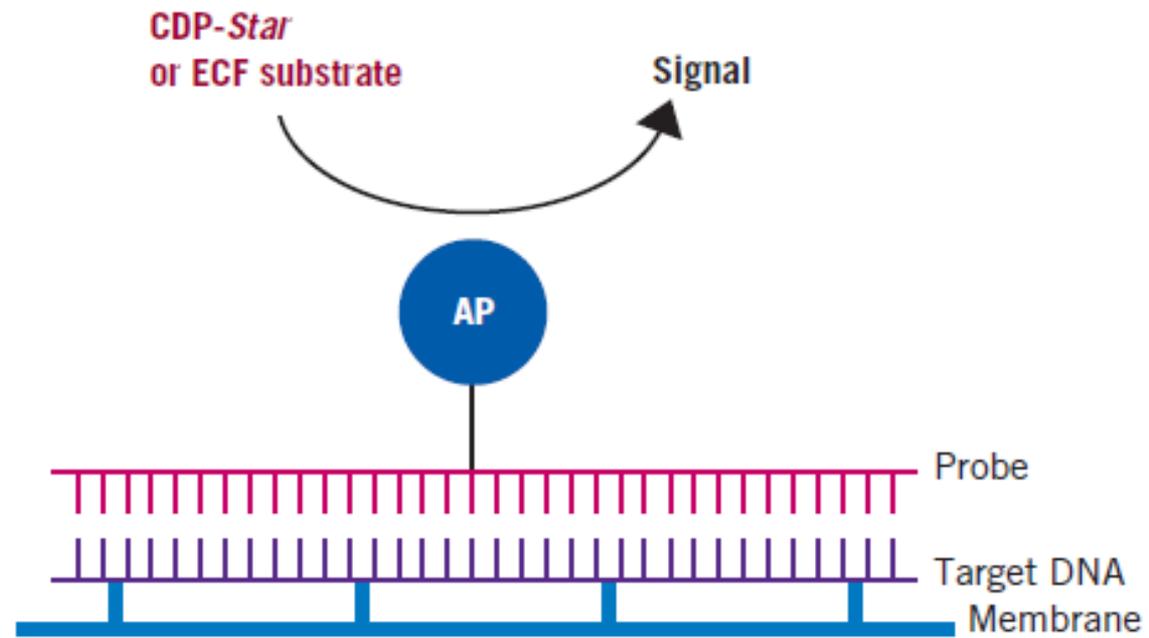


Fig 3. Outline of protocol for direct labelling method.

Amersham
ECL™ Direct Nucleic Acid
Labeling And Detection
Systems

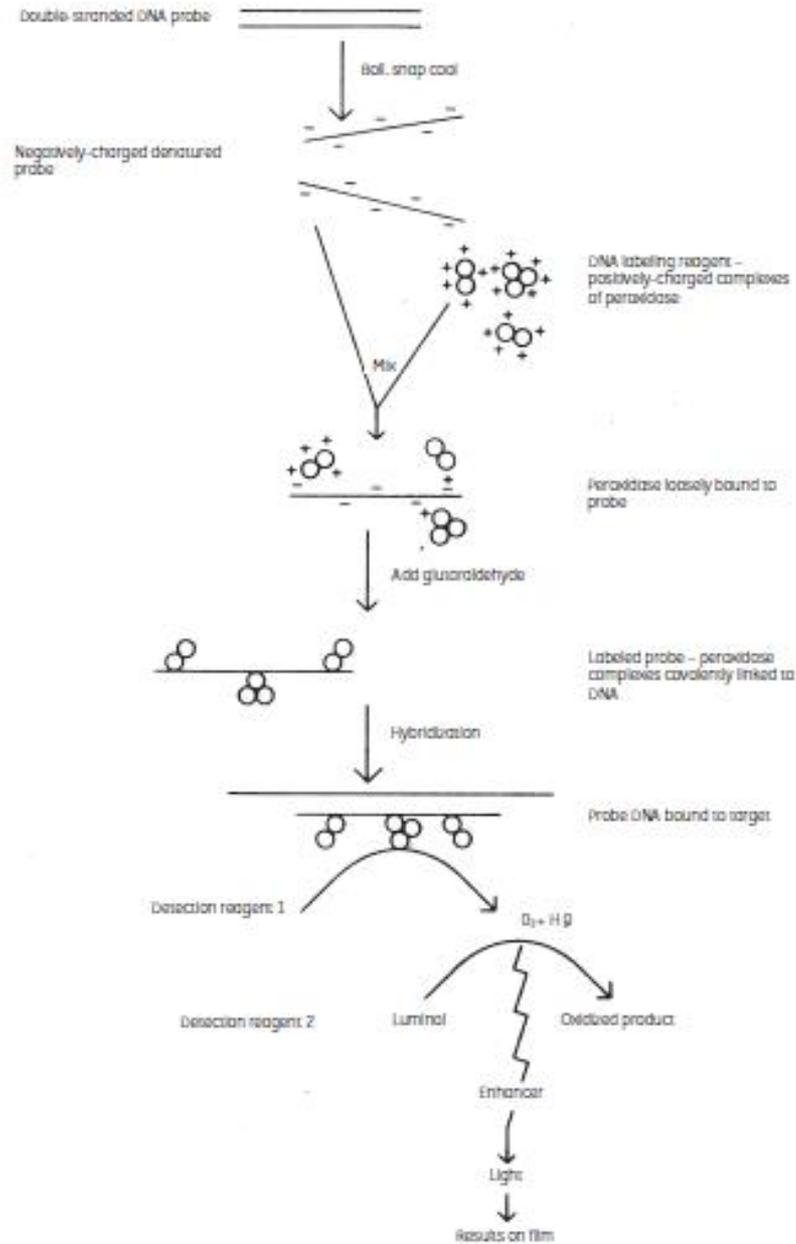
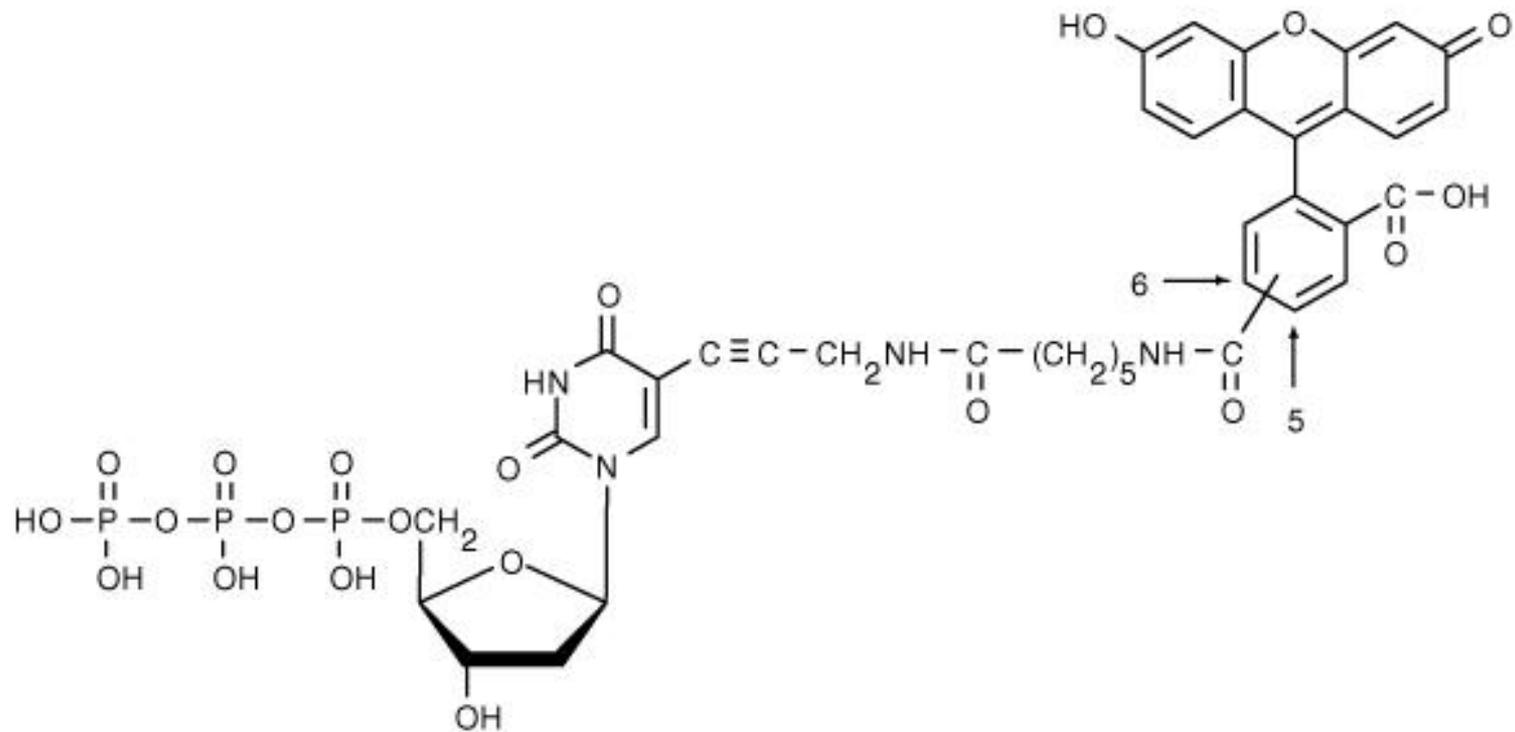
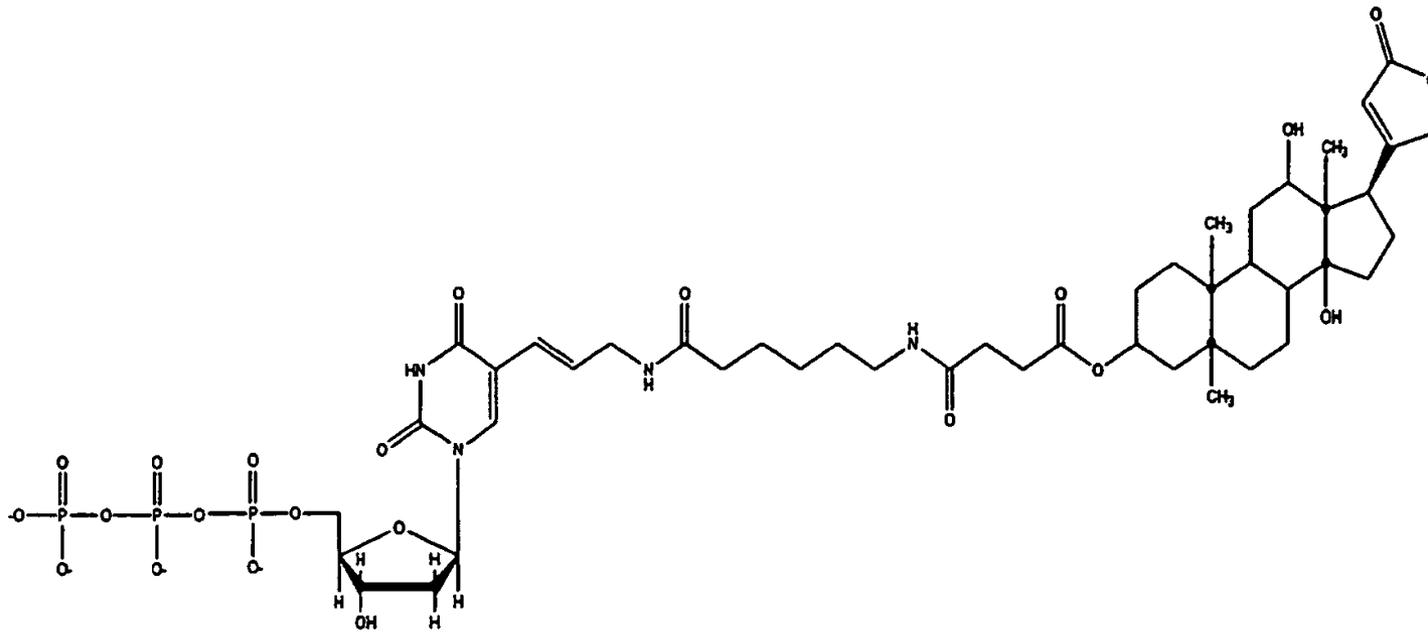


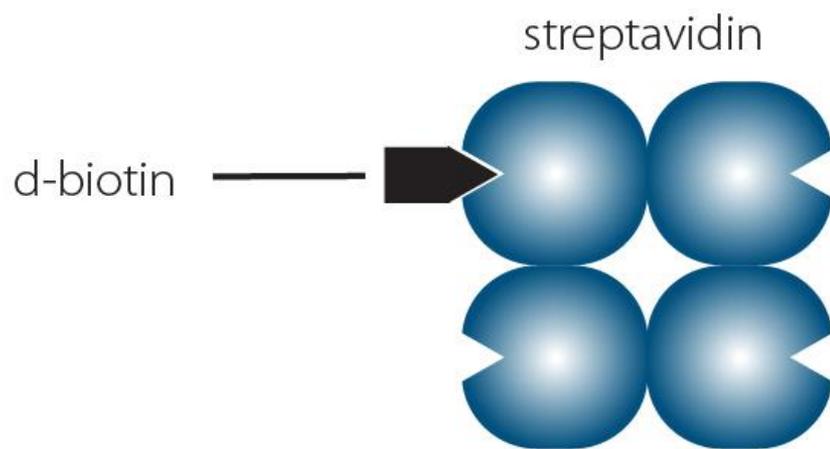
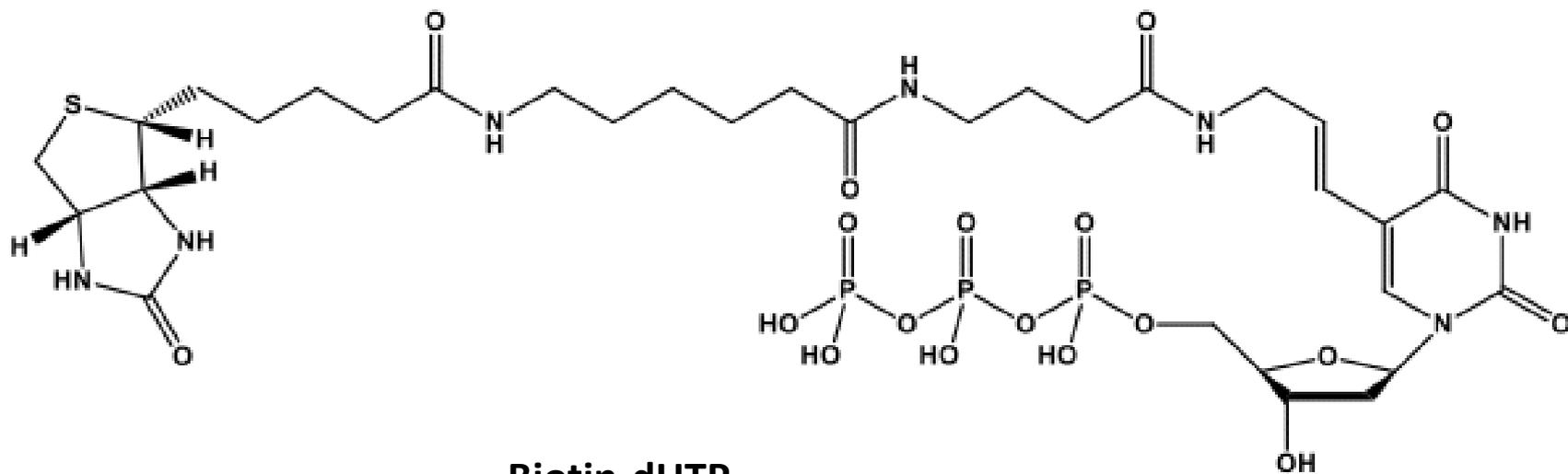
Figure 1. Principles of the ECL direct nucleic acid labeling and detection system.



Fluoresceína dUTP



Digoxigenina dUTP



$$K_d \approx 10^{-14} \text{ mol/L}$$

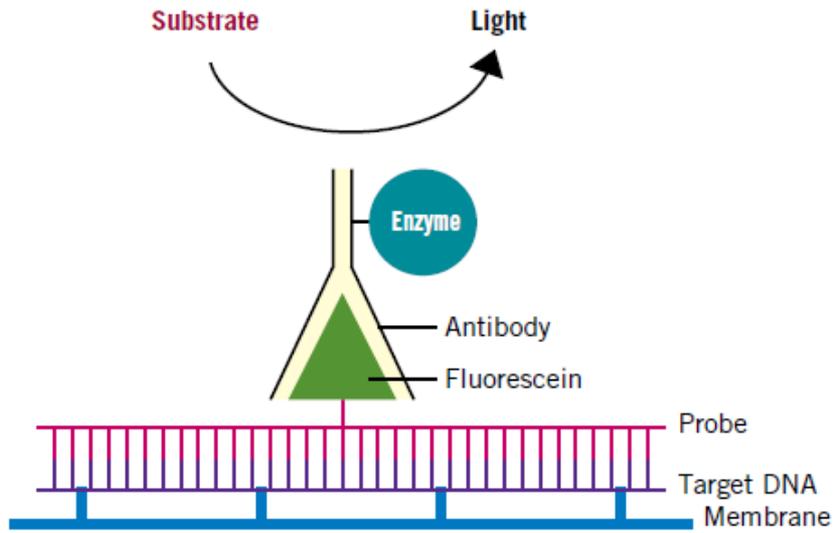


Fig 2. Outline of indirect labelling method.

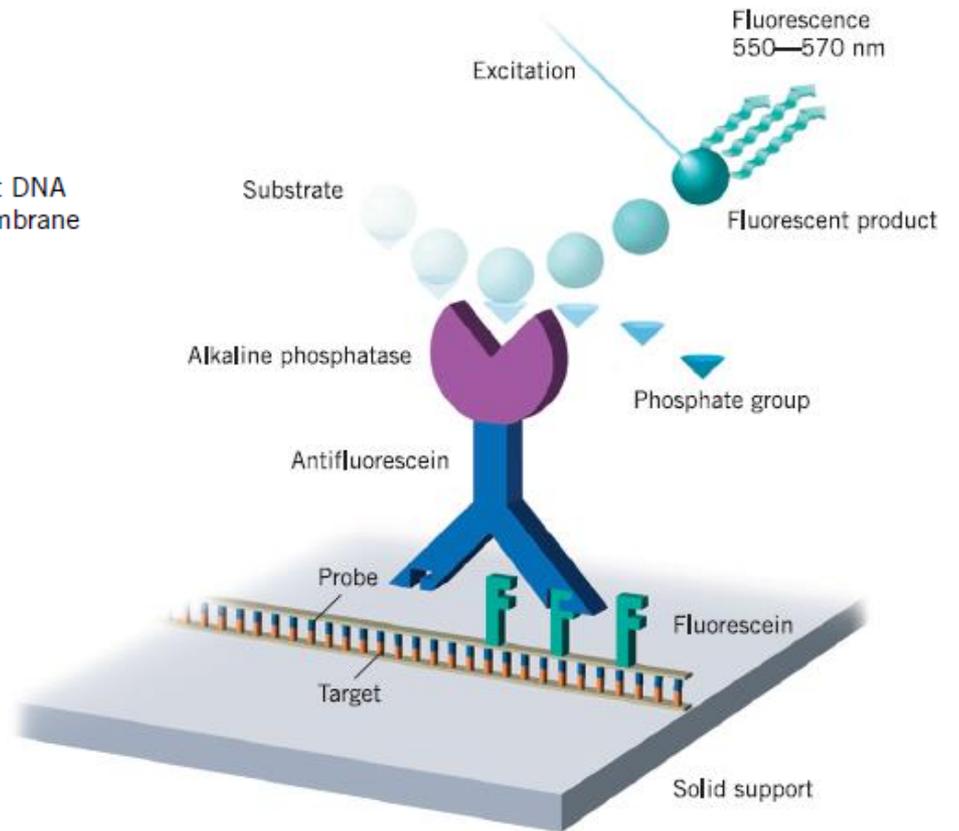
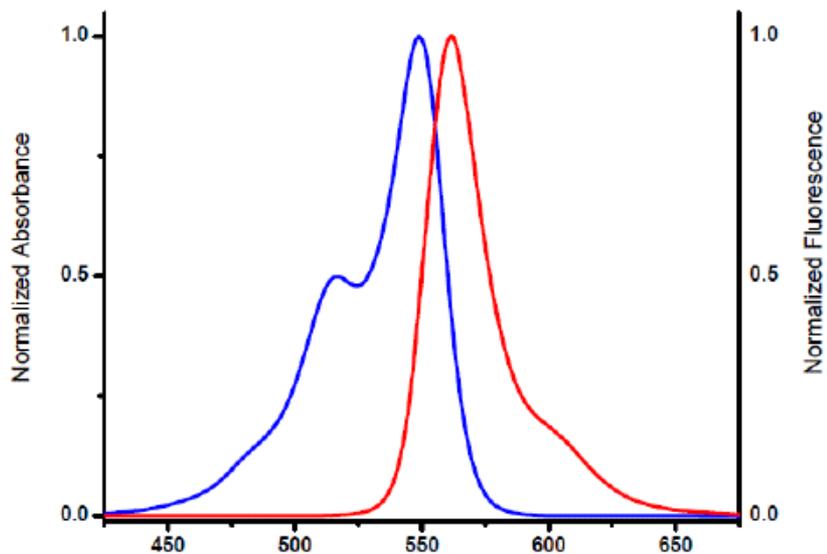


Table 1. Comparison of radioactive and non-radioactive nucleic acid labelling and detection systems.

Labelling and detection system	Sensitivity	Application	Probe labelling time	Time from hybridization to detection
³² P	Down to 10 fg	All high sensitivity applications	5 min to 3 h	On film 1 h to 1 week
AlkPhos Direct	60 fg	All high sensitivity applications	30 min	1 h
ECL Direct	0.5 pg	High target applications	20 min	1–2 h
Gene Images Random-Prime with CDP- <i>Star</i> Detection	50 fg	High sensitivity Northern blots	30 min	3 h
Gene Images 3'-Oligolabelling with CDP- <i>Star</i>	0.1 pg	Oligonucleotide screening with stringency control	30 min	3 h
Gene Images Random-Prime with ECF	0.25 pg	Quantification	30 min	3 h
Gene Images 3'-Oligolabelling with ECF	120 pg	Quantification	30 min	3 h
ECL Random-Prime	0.5 pg	Medium target Southern blots with DNA probes	30 min	3h
ECL 3'-End Labelling	0.2 pg	Medium to high target Southern blots with oligonucleotide probes	30 min	3 h



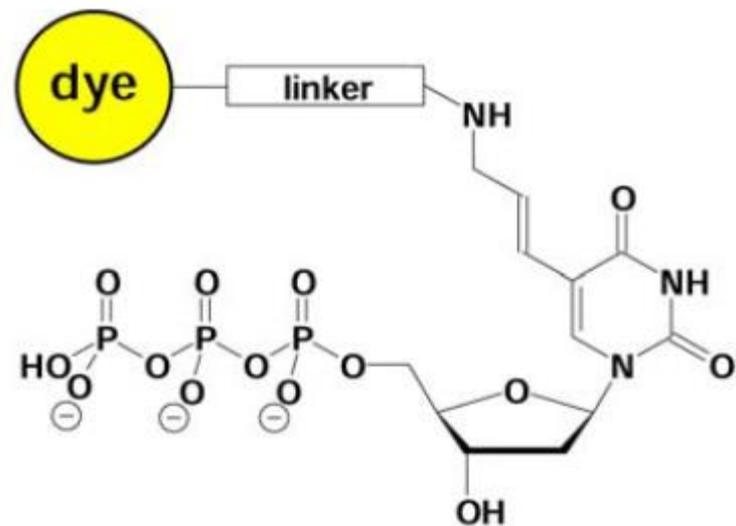
Cy3 excitation and emission spectra.

Spectroscopic data

Excitation maximum: $\lambda_{Ex} = 550 \text{ nm}$

Emission maximum: $\lambda_{Em} = 570 \text{ nm}$

Structure



Cy3-dUTP, the dye is attached via an optimized linker to aminoallyl-dUTP.

Molecular Structure:

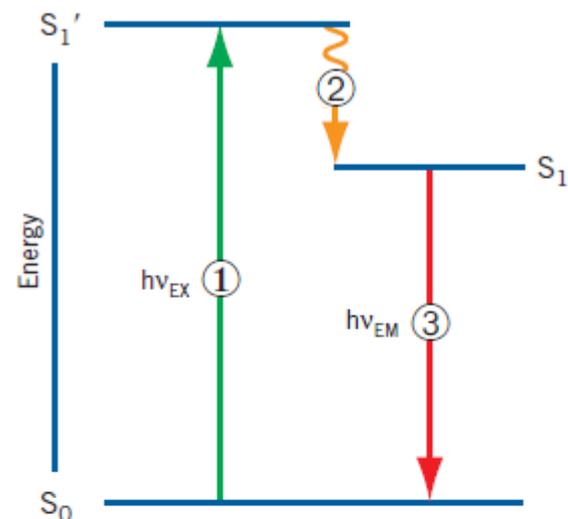
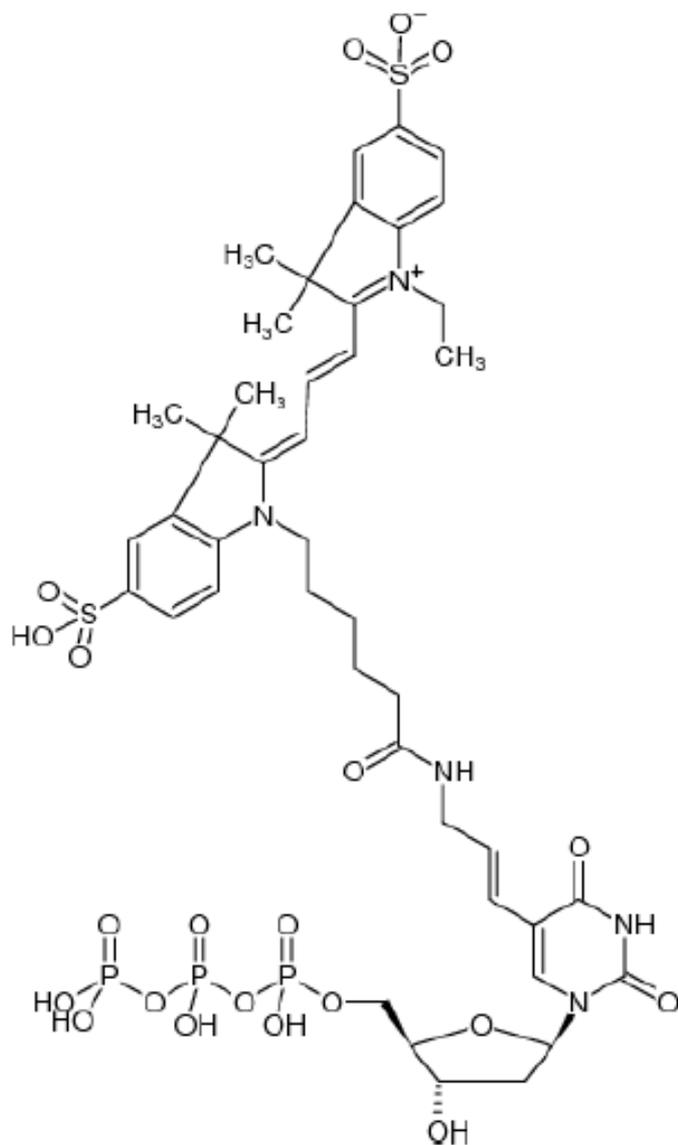
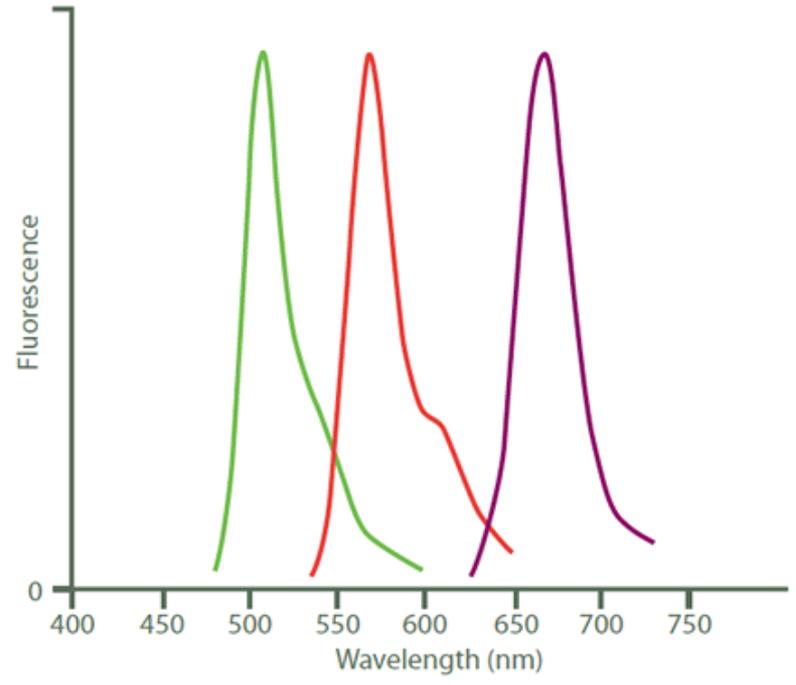
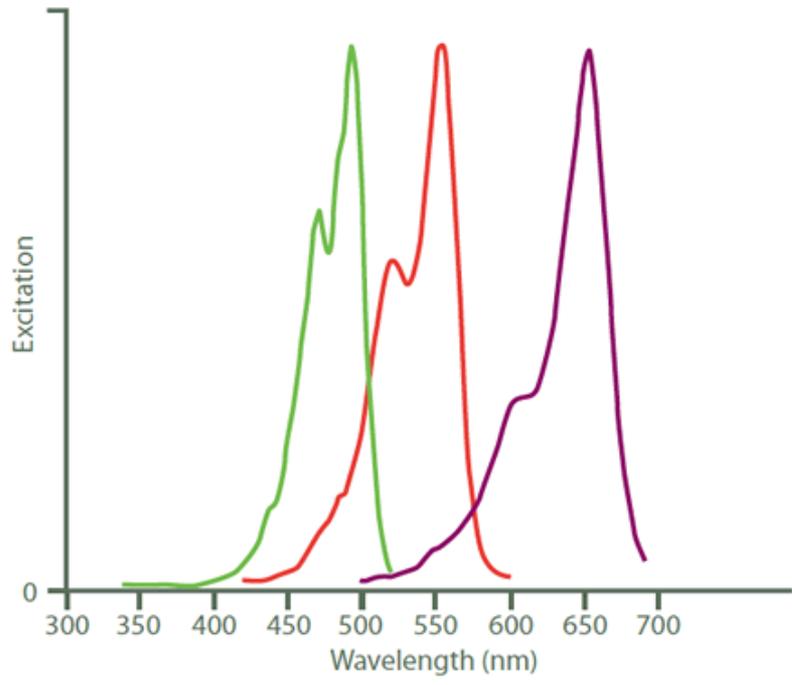


Fig 2. Jablonski diagram illustrating the processes involved in creating an excited electronic singlet state by optical absorption and subsequent emission of fluorescence. ① Excitation; ② Vibrational relaxation; ③ Emission.



Recommended PCR assay

20 μ l PCR labeling assay

Component	Stock conc.	Amount	Final conc.
High yield buffer without MgCl ₂	10x	2 μ l	1x
MgCl ₂ stock solution	25 mM	1.6 μ l	2 mM
dATP	1 mM	2 μ l	100 μ M
dCTP	1 mM	2 μ l	100 μ M
dGTP	1 mM	2 μ l	100 μ M
dTTP	1 mM	1 μ l	50 μ M
Cy3-dUTP	1 mM	1 μ l ¹⁾	50 μ M ¹⁾
forward Primer	10 μ M	1 μ l	500 nM
reverse Primer	10 μ M	1 μ l	500 nM
Template DNA		0.1-10 ng	5-500 pg/ μ l
Taq Pol	5 units/ μ l	0.2 μ l (1 unit)	0.05 units/ μ l
PCR grade H ₂ O		Fill up to 20 μ l	

1) The optimal final concentration of the labeled nucleotide may vary depending on the application.

Typhoon™



Fig 1. Typhoon FLA 9500 is a high performance, versatile laser scanner for sensitive and quantitative measurements in a multiuser environment.

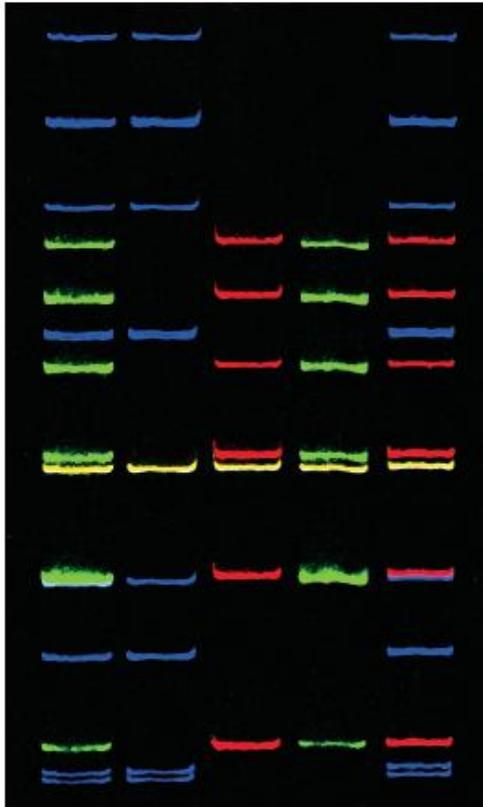


Fig 1. Fluorescently labelled DNA size ladders and PCR products loaded in the same lanes were electrophoretically separated in a polyacrylamide gel and imaged using Typhoon™ 8600 scanner. Fluorescein (green), CyTM3 (yellow), ROXTM (blue) and Cy5 (red) labels were used in amounts varying from 0.25 to 5 fmol per band.

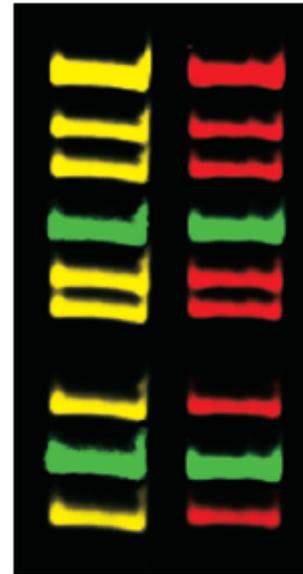


Fig 19. Three-colour gel image of a DNA in-lane sizing experiment. The fluorochromes used were TAMRATM (yellow), ROX (red), and fluorescein (green). The ROX and TAMRA bands are labelled DNA size ladders. The fluorescein fragments are PCR products of unknown size. Typhoon 8600 was used for image acquisition.

Microarrays

Se depositan decenas de miles de secuencias de DNA en una superficie pequeña, generalmente de vidrio (DNA chip). Las técnicas de *Northern blot* o qRT-PCR permiten estudiar unos pocos genes. Mediante *microarrays* se obtiene un perfil global de expresión.

Aplicaciones:

Cáncer: Clasificación de tumores, identificación genes supresores de tumores, biomarcadores, identificación de genes asociados con resistencia, *screening* de drogas etc

Asociación de genes con enfermedad.

Identificación de microorganismos patógenos, resistencia a antibióticos etc.

Genes expresados diferencialmente.

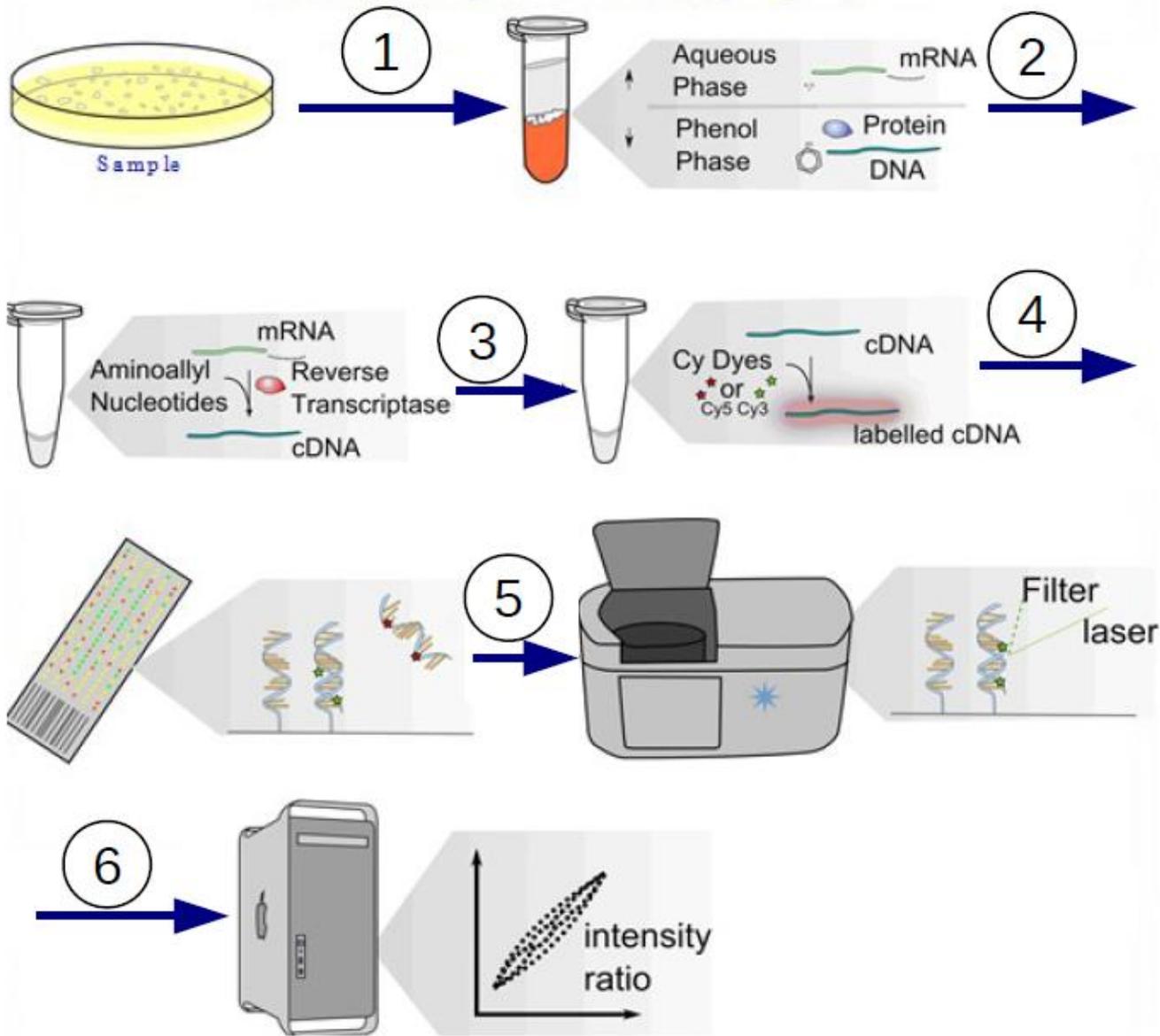
Limitaciones:

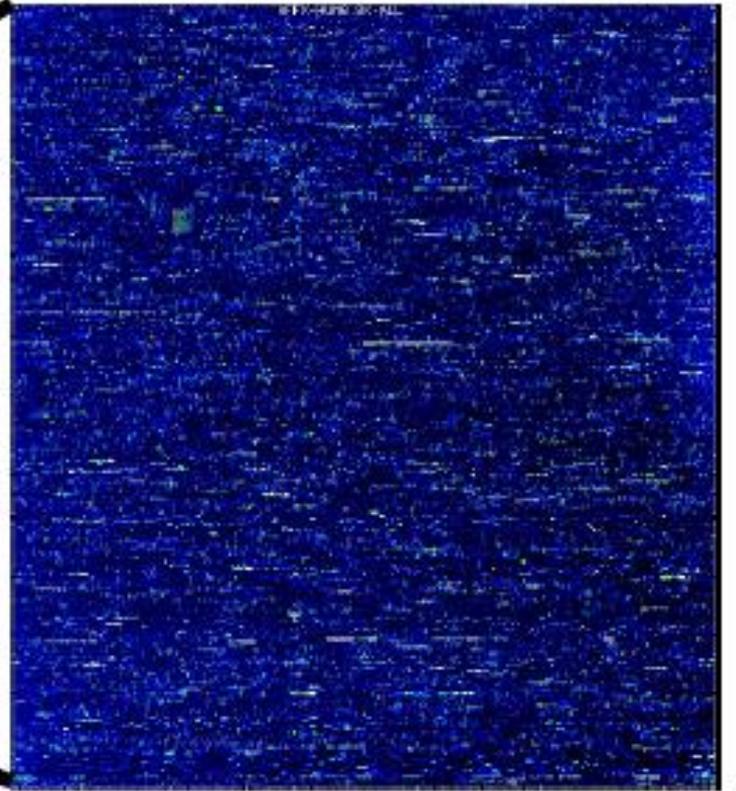
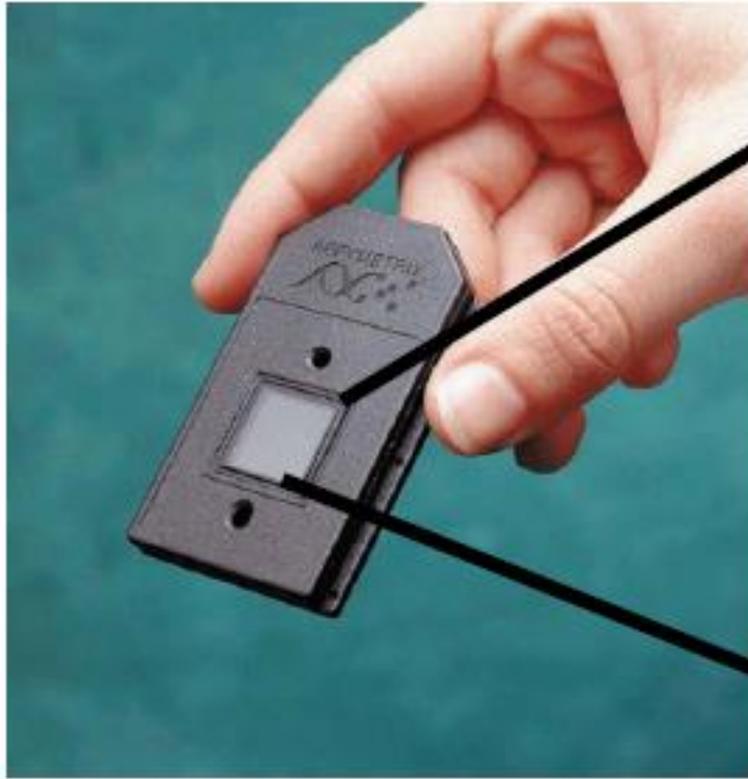
Solamente se detectarán secuencias para las cuales el *array* fue diseñado para detectar.

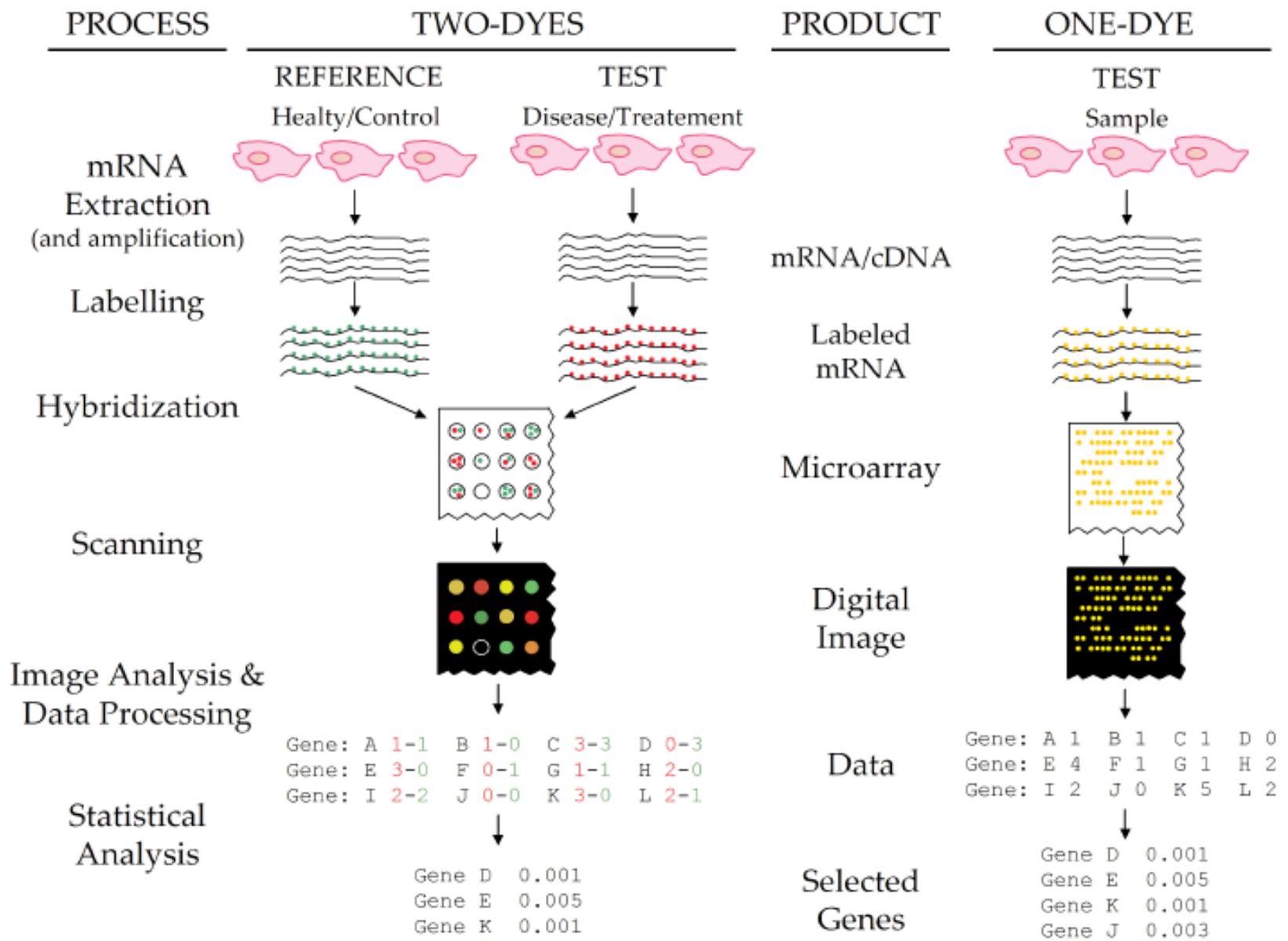
Muchas isoformas de cada gen.

La cuantificación es indirecta.

Especificidad.

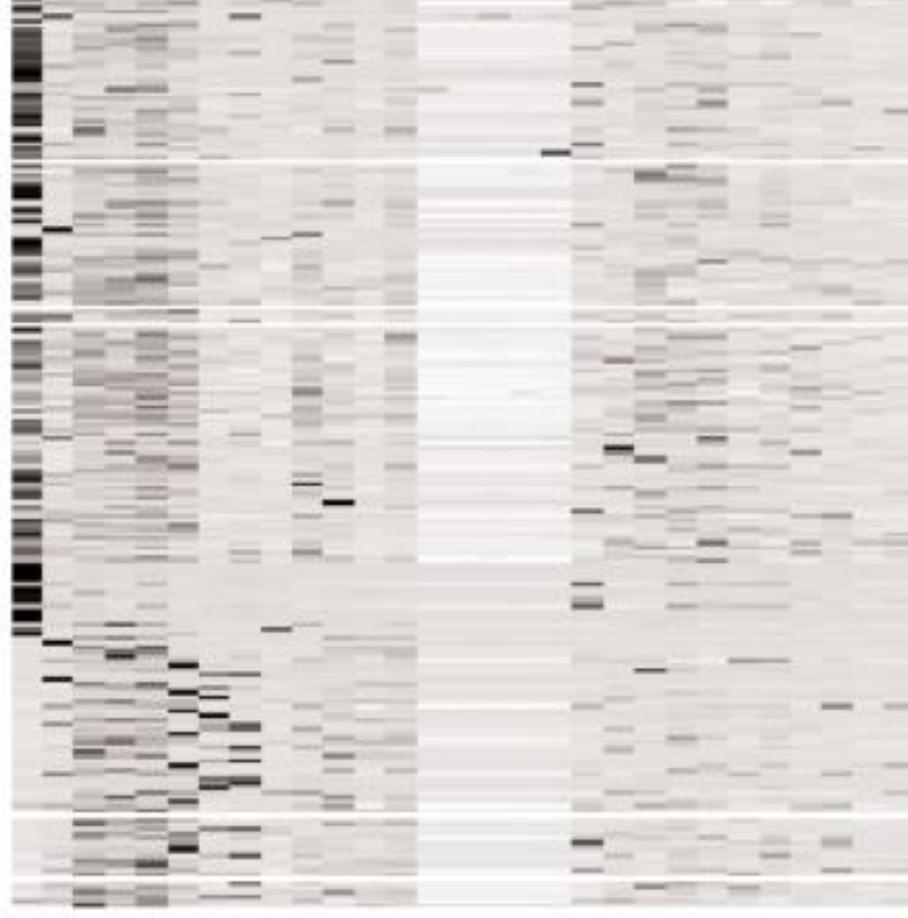






Placenta

Lung
Thalamus
Amygdala
Spinal Cord
Testis
Kidney
Liver
Pituitary
Thyroid
Cerebellum
Hypothalamus
Caudate Nucleus
Exocrine Pancreas
Lymph Node
Frontal Cortex
Stomach
Breast
Bone Marrow
Pancreatic Islets
Uterus
Ovary
Skin
Heart
Skeletal Muscle
Prostate
Thymus
Salivary Gland
Trachea



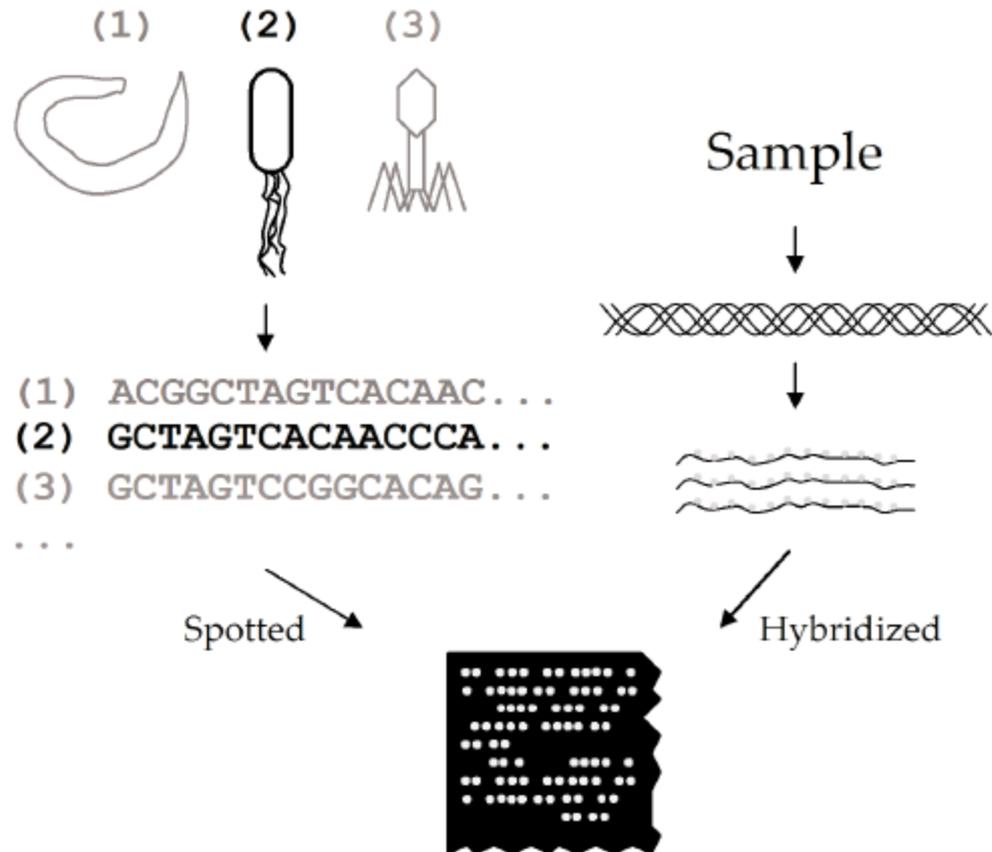


Figure 10. Multi-Pathogen Detection Using DNA Microarrays. Specific DNA sequences from disease-causing micro-organisms can be spotted on a microarray for pathogen detection.

Identification of microRNAs with Dysregulated Expression in Status Epilepticus Induced Epileptogenesis

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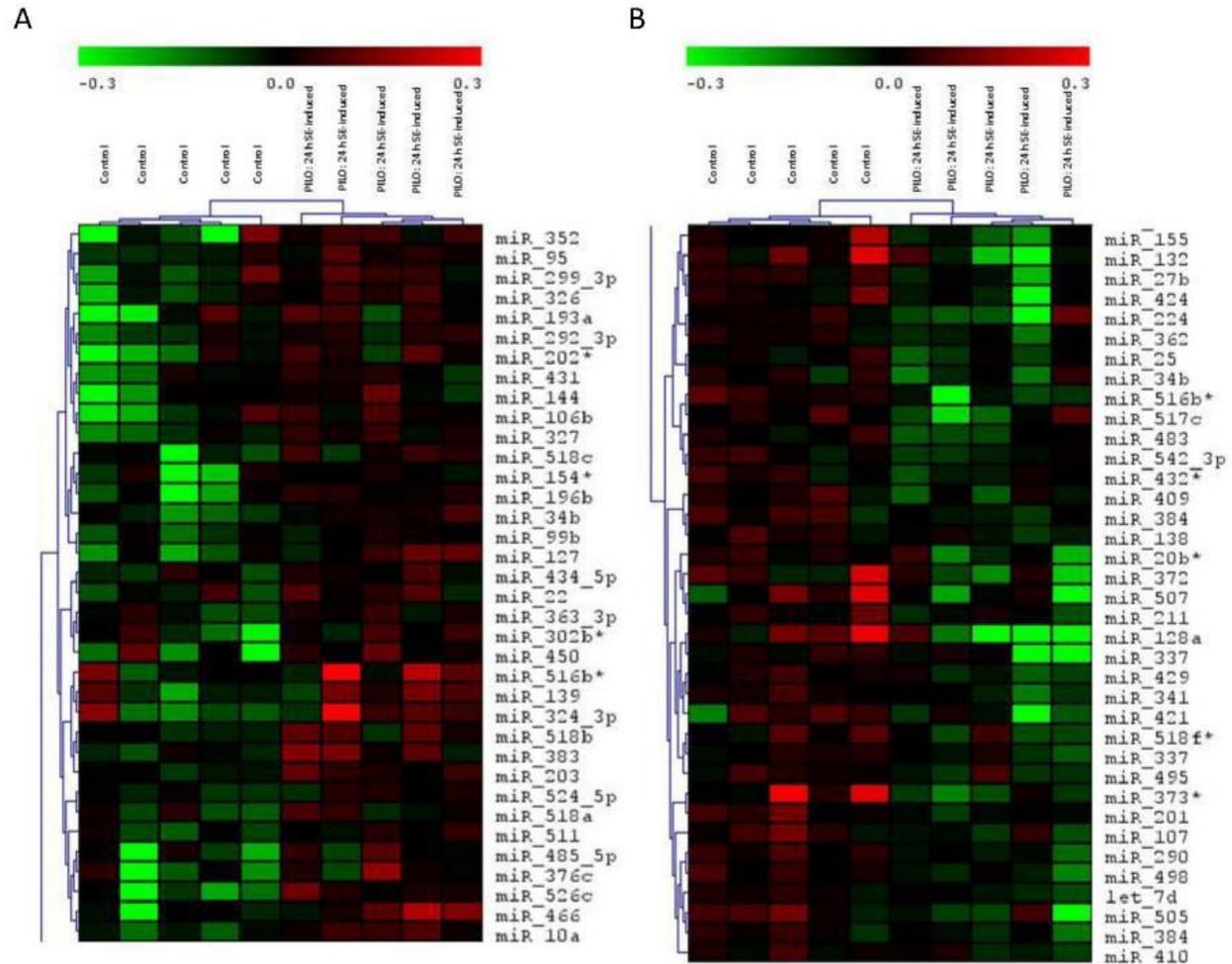


Fig 1. Hierarchical clustering of the 73 miRNA with significantly different expression in 24h post-SE hippocampus versus control experiment. Rows represent individual genes, and columns represent individual samples. The colorgram depicts high (red), average (black) and low (green) expression levels. A) Upregulated microRNAs. B) Downregulated microRNAs. (n = 5 for both H-PILO and control groups).